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Replacement-Free Mass-Amplified Sandwich Assay with 180-MHz Electrodeless Quartz-Crystal Microbalance Biosensor

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Abstract

This study presents the sensitivity-amplified detection method in the replacement-free electrodeless quartz crystal microbalance (QCM) biosensor. A sandwich assay is proposed for detecting C reactive proteins (CRP), where the biotinated second anti-CRP antibody is weighted by streptavidin for the sensitivity amplification. Because the first CRP antibody was immobilized nonspecifically on naked quartz surfaces, the sandwich assay was repeated using the same sensor chip, making the replacement-free assay possible. The mass-amplified sandwich assay detected the CRP solution of 0.1 ng/ml. A methodology for determining the molecular mass of the injected protein is also proposed.

Key words: Electrodeless QCM, Replacement Free, Sensitivity Amplification, CRP

1 Introduction

The quartz crystal microbalance (QCM) biosensor is mass-sensitive biosensor, and it allows detection of biomolecules captured by the corresponding receptor proteins immobilized on the oscillator surface through the change in the resonance frequency. The resonance-frequency change is principally caused by the mass loading effect (Sauerbrey (1959)). The viscosity effect (Kanazawa and Gordon (1985); Martin *et al.* (1991)) and the water-mass effect (Höök *et al.* (1998); Reimhult *et al.* (2004)) contribute to lower the resonance frequency as well, but their influence becomes insignificant at high frequencies (Ogi *et al.* (2009a)). The mass sensitivity of QCM biosensor improves as the fundamental resonance frequency increases, because high-frequency QCMs use thinner quartz plate, enhancing the captured mass. Therefore, developing a higher-frequency QCM biosensor remains central issue in the biosensor field (Uttenthaler *et al.* (2001); Cooper and Singleton (2007); Natesan *et al.* (2009); Furusawa *et al.* (2009)). We have proposed a wireless-electrodeless (WE) technique using an isolated quartz crystal and increased the fundamental frequency up to 170 MHz (Ogi *et al.* (2009a)).

As an important practical use, QCM biosensors are expected to detect biomarkers. However, human serum usually contains many blood proteins, and they are adsorbed on the sensor chip surfaces nonspecifically, making it difficult to extract the tiny frequency change caused by the binding of a target protein with the receptor on the surface from the total frequency response. Also, threshold concentrations are usually smaller than the detection limit of QCM biosensors. For overcoming this difficulty, we propose a mass-amplified sandwich assay using biotinized antibody, whose mass can be increased effectively by attaching

streptavidin owing to extremely high affinity between biotin and streptavidin. As a target protein, we select the C-reactive protein (CRP), which is an important biomarker for checking inflammation. Its threshold level is about 30 ng/ml, and very quick assay is often required in an emergency case to judge the cause of illness. Thus, the WE-QCM can be a candidate because of its reusability and high sensitivity.

In the previous studies, gold nano particles have been used for the mass amplification in QCM assays (Mao *et al.* (2006); Zhu *et al.* (2009); Chen *et al.* (2010); Pinto *et al.* (2010)). However, high affinity of gold with proteins (Ogi *et al.* (2009b)) causes nonspecific adsorption on receptor proteins. Also, preparation of surface modified particles takes longer time. Instead of the gold nano particles, we propose the mass amplification method using streptavidin, which quickly combines with biotin with extremely high affinity. Our mass-amplified sandwich assay detects CPR with concentration of 100 pg/ml. Furthermore, we propose a useful methodology to determine molecular mass of an injected ligand and apply it to the mass-amplified second anitbody.

2 Experimental Section

The first and second anti-CRP antibodies were obtained from Bethyl Laboratories, Inc. (No. A80-125A and No. A80-125F, respectively). CRP was obtained from Acris Antibodies GmbH (No. BA1087). The NH₂-reactive biotin was from Dojindo (biotin labeling Kit-NH₂). Streptavidin was from IBA Headquarters (No. 2-0203-001).

The proposed assay takes three steps. First, the first anti-CRP antibody is

immobilized on the quartz surfaces nonspecifically. Second, the analyte (CRP solution) is injected. Third, the mass-amplified second anti-CRP-antibody solution is injected after the injection of the carrier solution for washing the surfaces. The schematic diagram of the sandwich assay is shown in Fig. 1(a).

A rectangular 9- μm thick AT-cut bare quartz ($2.5\text{ mm} \times 1.7\text{ mm}$), whose fundamental resonance frequency f is near 182 MHz, was located at middle part of the flow channel by lightly sandwiching its corner edge by 1-mm thick silicon-rubber gaskets. The carrier solution was phosphate buffered saline (PBS) solution (pH 7.4), and nearly entire both surfaces of the quartz plate were exposed to the flowing solution. The two straight line antennas and a copper foil for grounding were embedded in bottom and top covers, respectively (Ogi *et al.* (2009a)). The volume of the flow channel in the sensor cell is $2 \times 3 \times 40\text{ mm}^3$. The flow-injection analysis system used was homebuilt (Ogi *et al.* (2007)).

The tone burst signal was applied to the generation antenna to excite the shear vibration, and the reverberating signals of the quartz plate were received by the detection antenna with noncontacting, which were processed by a super-heterodyne phase sensitive detector. The resonance frequency of the quartz was determined from the phase of the received signal. Details of the signal processing can be found elsewhere (Ogi *et al.* (2009c)).

The quartz crystal was first cleaned in a piranha solution ($98\% \text{ H}_2\text{SO}_4:33\% \text{ H}_2\text{O}_2=7:3$). After rinsing with ultrapure water, it was immersed in $500\text{ }\mu\text{g/ml}$ first anti-CRP-antibody solution for 5 h at $4\text{ }^\circ\text{C}$ and rinsed by the PBS solution. Because the naked quartz surface adsorbs immunoglobulin G (Ogi *et al.* (2009b)), it is possible to immobilize the first antibody just by immersing the cleaned sensor chip in the antibody solution.

We prepared the mass-amplified second anti-CRP antibody as follows: The second antibody dissolved in the buffer solution was centrifuged in the filtration tube for 10 min with 8000g for removing smaller foreign substances. The NH_2 -reactive biotin (biotin with active ester group at its end) was dissolved with 10 μl dimethyl sulfoxide (DMSO) and poured into the filtration tube together with a 100 μl reaction buffer and incubated for 30 min at 37°C . Then, the buffer solution was added in the tube and the solution was centrifuged to remove unreacted biotin molecules. This procedure was repeated three times, and we obtained the biotinated second anti-CRP antibody solution. The possible maximum concentration of the solution is 1 $\mu\text{g}/\text{ml}$ estimated from the concentration of initial antibody solution, but this value could be lower because some second-antibody molecules were taken in the filter and uncollected.

Then, we mixed streptavidin in the second anti-CRP antibody solution, where concentrations of streptavidin was 1 $\mu\text{g}/\text{ml}$. This solution was incubated for 30 min at 37°C before the injection.

CRP solutions (0.1, 1, 10, and 100 ng/ml , in PBS) were prepared and injected in the reaction cell containing first antibody adsorbed on the quartz plate. The PBS solution was injected for a while for washing the sensor surfaces, and then the mass-amplified second antibody solution was injected.

3 Results

Figure 2 shows an example of the enhanced frequency change by the mass-amplified second-antibody solution. Injection of the 1 ng/ml CRP solution caused only 200-Hz frequency decrease, and the baseline increased after 15

min from the injection. Thus, it would have been difficult to estimate the amount of the frequency change due to the binding of CRP with first antibody. However, the injection of the mass-amplified second-antibody solution caused nearly 5-kHz frequency change, demonstrating the successful amplification of the mass sensitivity.

Figure 3(a) shows the binding curves observed after the arrival of the mass-amplified second-antibody solution, which were followed by injection of CRP solutions and the PBS solution. We observed the frequency decrease even for the injection of the second-antibody solution after the analyte solution without CRP molecules (0 ng/ml), indicating the nonspecific binding of the second antibody onto the first antibody. However, this frequency change was significantly smaller than those observed for the analyte solutions containing CRP molecules. The inset in Fig. 3(a) shows the correlation between the frequency decrement at 20 min from the arrival time of the mass-amplified second-antibody solution and the concentration of the CRP solution. However, such a correlation was obtained when we used antibodies from the same production lots, and the frequency change fluctuated among the assay with those from different lots as shown in Fig. 3(b).

4 Discussion

First, we emphasize that the electrodeless QCM is the replacement-free biosensor and the sensor chip was reusable after an assay by washing it with the piranha solution and rinsing with ultrapure water. This is an important advantage in treating many analyte solutions because and is only achieved by the electrodeless technique. The assay time (~ 20 min from the injection of the

mass amplified solution) is shorter than those in conventional enzyme-linked immunosorbent assays ($>\sim 3$ h).

Figure 2 clearly demonstrates that the mass-amplified second antibody increases the amount of the frequency change. We here propose a useful method to estimate molecular mass of injected ligand from a single frequency-monitoring measurement such as Fig. 2. This is made possible with the benefit of the high-frequency WE-QCM because of its high sensitivity to the added mass added on the sensor surfaces. Consider an equilibrium binding reaction between proteins A and B :



Where A and B denote the injected protein and the receptor protein on the quartz surfaces, respectively, and AB denotes their complex on the surface (Figure 1(a) illustrates the case between the mass amplified antibody for A and the CRP on the surface for B .) Assuming the pseudo-first-order reaction, the frequency change during the binding reaction ($A + B \longrightarrow AB$) and that during the dissociating reaction ($A + B \longleftarrow AB$) obey exponential functions with exponential coefficients α_1 and α_2 , respectively (Liu *et al.* (2003)). (In Fig. 2, we showed fitted exponential functions to extract the coefficients with red lines.) These coefficients relate with the reaction-velocity constants as

$$\alpha_1 = C_A k_a + k_d, \quad (2)$$

$$\alpha_2 = k_d. \quad (3)$$

Here, we denote k_a and k_d as the reaction velocity constants for association and dissociation reactions, respectively, and C_A denotes the bulk concentration of molecule A . Therefore, the equilibrium constant $K_A = k_a/k_d$ can be

determined by

$$K_A = \frac{1}{C_A} \left(\frac{\alpha_1}{\alpha_2} - 1 \right). \quad (4)$$

At the equilibrium state, the area molecular density of the complex AB , C_{AB} , can be expressed by (Ogi *et al.* (2009b))

$$C_{AB} = \frac{K_A C_A C_B^0}{K_A C_A + 1}. \quad (5)$$

C_B^0 denotes the area molecular density of the receptor protein on the surface, and based on the Sauerbrey equation, it can be expressed by the frequency change Δf_B observed when the receptor protein B is injected (see Fig. 1(b)) as

$$C_B^0 = \frac{1}{2} \frac{\rho_q t_q \Delta f_B}{M_w^B f} \quad (6)$$

Where ρ_q and t_q are mass density and thickness of the quartz plate, respectively, and M_w^B denotes the molecular mass of protein B . The factor $\frac{1}{2}$ appears because both surfaces can be the sensing region in our QCM. Considering that C_{AB} is equal to the area molecular density of captured protein A on surfaces, and it can be obtained from the frequency change Δf_A like as C_B^0 , we can determine the molecular mass of protein A , M_w^A from equations (4) to (6) as

$$M_w^A = M_w^B \left(\frac{\Delta f_A}{\Delta f_B} \right) \left(1 - \frac{\alpha_2}{\alpha_1} \right)^{-1} \quad (7)$$

This is a useful equation because we can determine the molecular mass of the injected protein from a single frequency measurement, and this can be made possible when the frequency of the QCM is so high that the simple Sauerbrey equation applies. We confirmed this equation using staphylococcus aureus protein A as the receptor protein (B) and human immunoglobulin G

(hIgG) as the analyte (A), and obtained the predicted molecular mass of hIgG between 143 and 160 kDa, close to the molecular mass of hIgG of 150 kDa. (Because we published many QCM measurements on this binding system (Ogi *et al.* (2009a, 2007, 2008)), we omit showing these experimental data.)

Using this theory, we estimate the molecular mass of the mass amplified second anti-CRP antibody. For example, we observe $\Delta f_B = \sim 200$ Hz with the injection of the 1 ng/ml CPR solution and $\Delta f_A = \sim 4.6$ kHz caused by injection of the mass-amplified second-antibody solution in Fig. 2, yielding the molecular mass of the mass-amplified second antibody to be about 3,200 kDa with the molecular mass of CPR of 105 kDa. This value is much larger than the molecular mass of the unmodified second antibody (~ 150 kDa) by a factor of 20. Therefore, mass amplification can be enhanced significantly. Because the IgG based antibody possesses several amino group terminals, a single biotin-ated antibody can bind with several streptavidin molecules (molecular mass: 53 kDa). Furthermore, because a single streptavidin has four binding sites with biotin, it can connect more than two antibody molecules through biotin terminals. The molecular mass of 3,200 kDa is an averaged value, and we cannot identify the structure of the second-antibody complex. (It corresponds to a complex of six second antibodies with seven streptavidins attached to each antibody, for example.) The size will be affected by the incubation time and will affect the affinity between second antibody and CPR. Thus, it will be needed to control the incubation time in preparing the second-antibody solution to achieve the most suitable assay condition in our future study.

Finally, we discuss about the sensitivity on the CRP detection. We see a good correlation between the CRP concentration and the frequency decrement in Fig. 3(a), suggesting the applicability of the proposed sandwich method to di-

agnosis with marker proteins. Such a correlation was obtained when we used the substances from the same sample lots, but the fluctuation of the frequency response becomes worse among measurements for different sample lots (Fig. 3(b)). Even in this case, the proposed assay can detect a low concentration CRP solution (100 pg/ml), for which the direct assay without second antibody failed to detect, confirming the high sensitivity of this assay. In this study, we used polyclonal antibodies, and their binding affinity and epitopes on CRP usually depend on the production lot. Also, the preparation of the mass-amplified second antibody requires high-affinity binding reactions, and the reaction time and temperature should significantly affect size and affinity of the second antibody. Thus, stable productions of individual substances with monoclonal antibodies will be needed for reliable diagnosis, and we will investigate systematically the sensitivity and reproducibility of this assay by changing time and temperature in preparing the mass-amplified second antibody with monoclonal antibodies.

5 Conclusions

We proposed mass-sensitivity amplification method using purely protein substances for the wireless-electrodeless QCM with a fundamental resonance frequency of 182 MHz. The sandwich assay was performed for detecting CRP. The first antibody was first immobilized nonspecifically, and the mass-amplified second antibody was injected, following the injection of the CRP solution. The detection was possible even for a smaller concentration CRP solution with 0.1 ng/ml. From the binding curves and the presented theory, the molecular mass of the mass-amplified second antibody was estimated to be 3,200 kDa.

The frequency response was fluctuated among measurements with substances from different production lots, and further improvements for purification of substances with monoclonal antibodies and strict controlling on time and temperature on preparation of the mass-amplified second antibody is needed for a reliable diagnosis use, which will be achieved in near future.

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Figure Captions

Fig. 1 (Color)(a)Schematic of the sandwich assay using the mass-amplified second anti-CRP antibody. A, B, and AB denote the mass-amplified second antibody, the CRP adsorbed on the first antibody, and their complex, respectively. (Note that actual numbers of second antibody and streptavidin are larger than those in this schematic.) (b)Schematic of a frequency response during the immobilization, binding, and dissociating reactions.

Fig. 2 (Color)Frequency response for injections of a 1 ng/ml CRP, PBS, and the mass-amplified second-antibody solutions. Arrows indicate the arrival time of the solutions at the sensor cell.

Fig. 3 (a) Binding curves observed for the injection of the mass-amplified second-antibody solution after the injection of the CRP solution with various concentrations. Arrows indicates arrival of the PBS solution. The numbers denote the concentration of the CRP solution injected before the mass-amplified second-antibody solution. The inset shows the correlation between the frequency change and the CRP concentration C_{CRP} at 20 min. (b)Frequency decrement and CRP concentration C_{CRP} at 20 min for measurements with antibodies from different production lots.

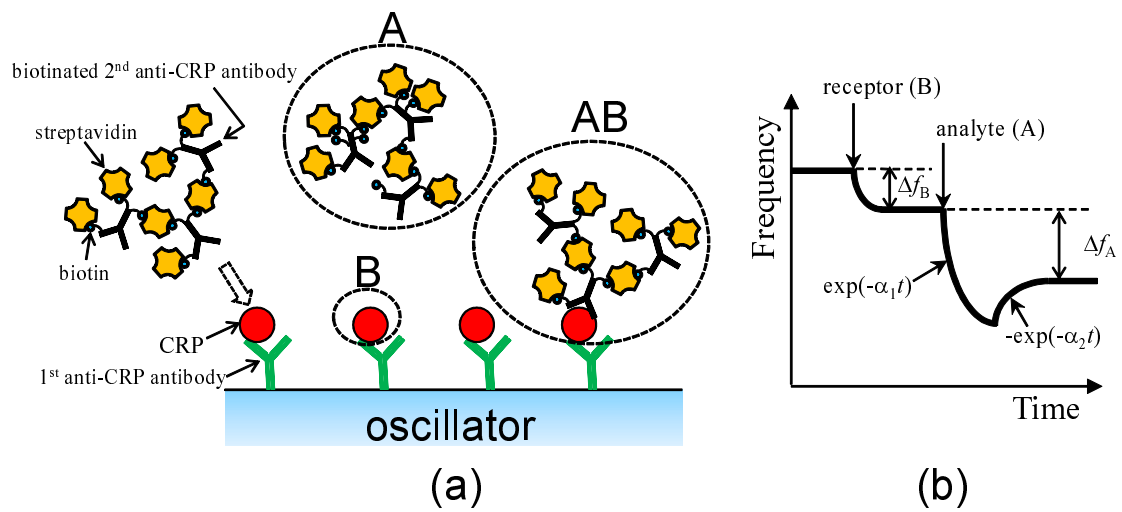


Fig. 1.

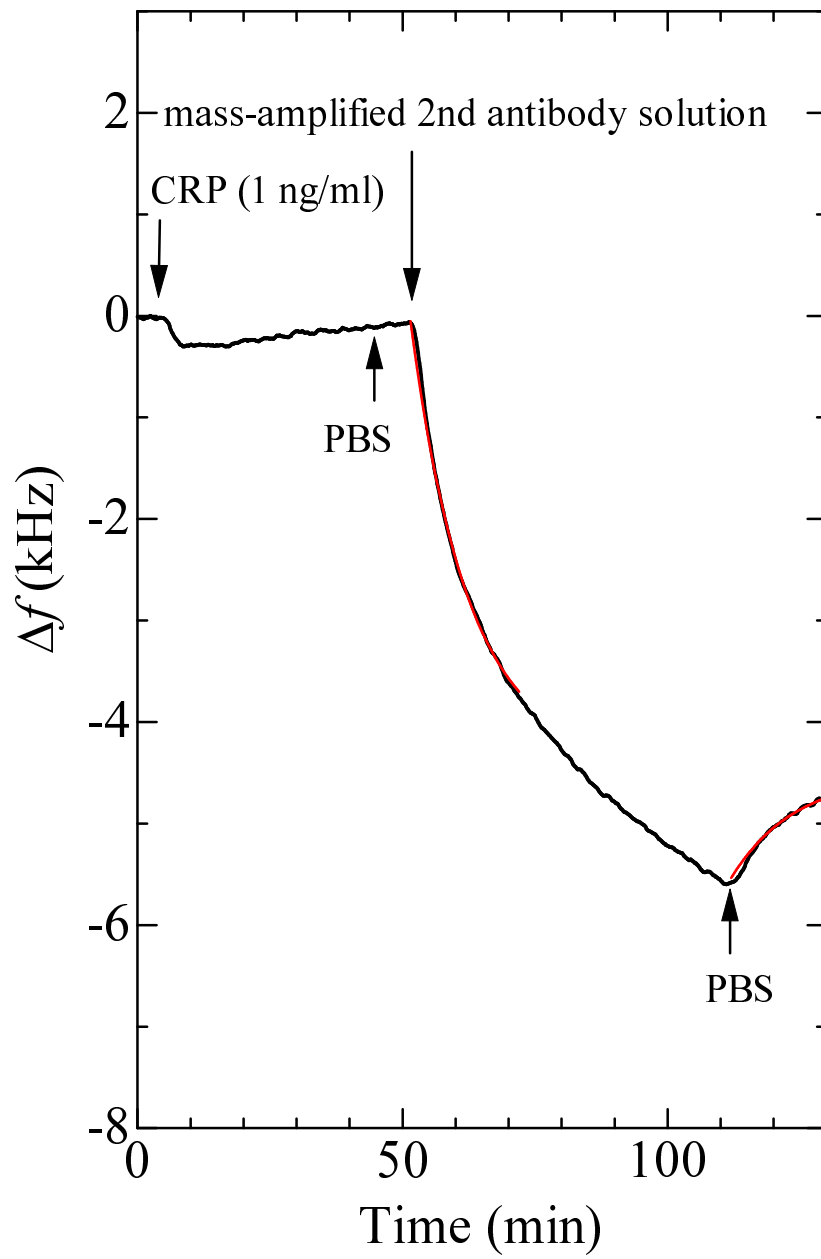


Fig. 2.

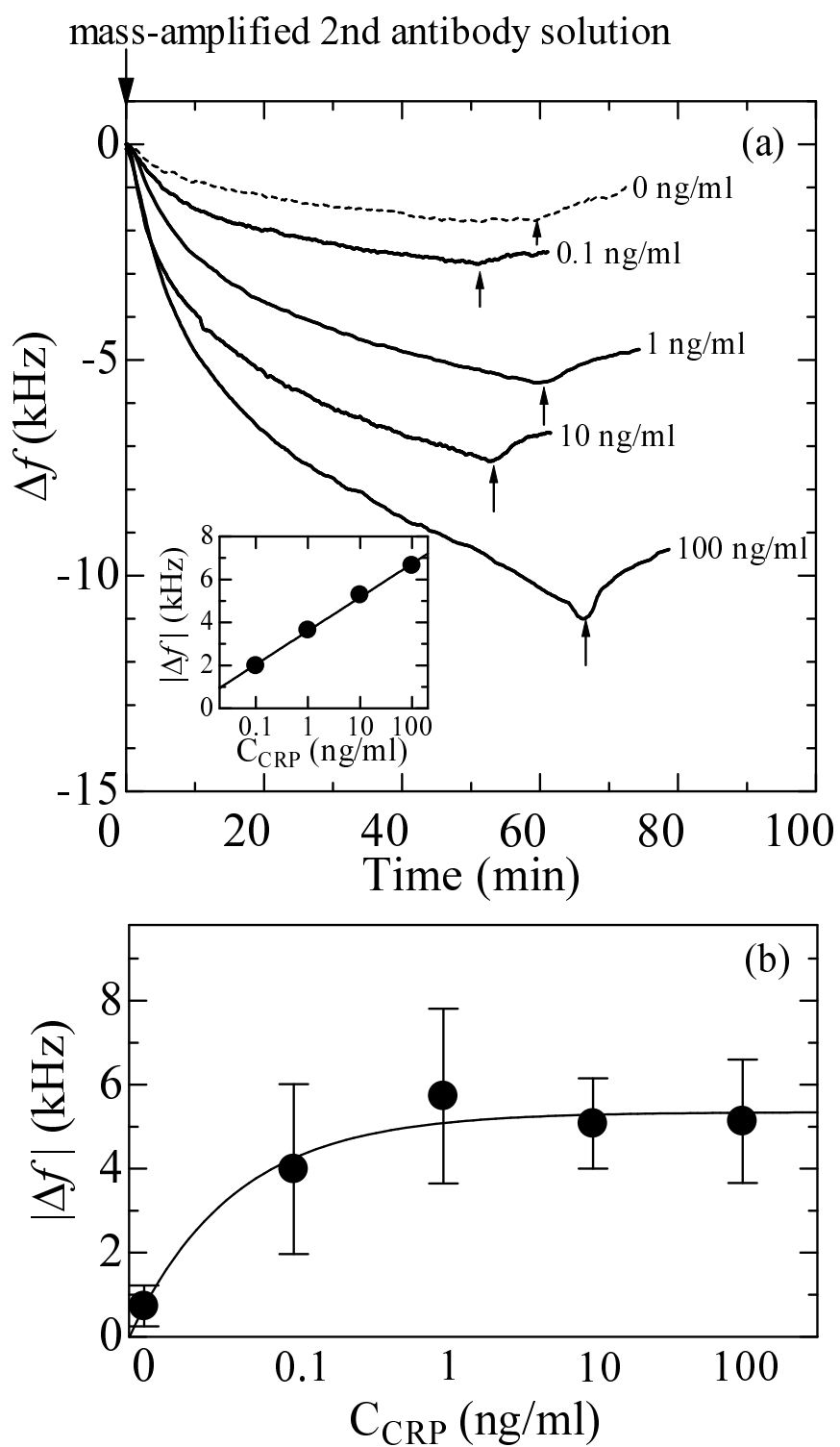


Fig. 3.