



Natural Product Research: Formerly Natural Product Letters

Publication details, including instructions for authors and subscription information:

<http://www.tandfonline.com/loi/gnpl20>

Anti-quorum sensing activity of essential oils from Colombian plants

Beatriz Jaramillo-Colorado ^{a b}, Jesus Olivero-Verbel ^a, Elena E. Stashenko ^c, Irene Wagner-Döbler ^d & Brigitte Kunze ^d

^a Environmental and Computational Chemistry Group, Faculty of Pharmaceutical Sciences, University of Cartagena, Cartagena, Colombia

^b Agrochemical Research Group, Chemistry Program, Faculty of Exact and Natural Sciences, University of Cartagena, Cartagena, Colombia

^c Chromatography Laboratory, Research Centre for Biomolecules, CIBIMOL, CENIVAM, Industrial University of Santander, Bucaramanga, Colombia

^d Helmholtz Centre for Infection Research, Group Microbial Communication, Braunschweig, Germany

Published online: 21 Sep 2011.

To cite this article: Beatriz Jaramillo-Colorado, Jesus Olivero-Verbel, Elena E. Stashenko, Irene Wagner-Döbler & Brigitte Kunze (2012) Anti-quorum sensing activity of essential oils from Colombian plants, Natural Product Research: Formerly Natural Product Letters, 26:12, 1075-1086, DOI: [10.1080/14786419.2011.557376](https://doi.org/10.1080/14786419.2011.557376)

To link to this article: <http://dx.doi.org/10.1080/14786419.2011.557376>

PLEASE SCROLL DOWN FOR ARTICLE

Taylor & Francis makes every effort to ensure the accuracy of all the information (the "Content") contained in the publications on our platform. However, Taylor & Francis, our agents, and our licensors make no representations or warranties whatsoever as to the accuracy, completeness, or suitability for any purpose of the Content. Any opinions and views expressed in this publication are the opinions and views of the authors, and are not the views of or endorsed by Taylor & Francis. The accuracy of the Content should not be relied upon and should be independently verified with primary sources of information. Taylor and Francis shall not be liable for any losses, actions, claims, proceedings, demands, costs, expenses, damages, and other liabilities whatsoever or

howsoever caused arising directly or indirectly in connection with, in relation to or arising out of the use of the Content.

This article may be used for research, teaching, and private study purposes. Any substantial or systematic reproduction, redistribution, reselling, loan, sub-licensing, systematic supply, or distribution in any form to anyone is expressly forbidden. Terms & Conditions of access and use can be found at <http://www.tandfonline.com/page/terms-and-conditions>

Anti-quorum sensing activity of essential oils from Colombian plants

Beatriz Jaramillo-Colorado^{ab}, Jesus Olivero-Verbel^{a*}, Elena E. Stashenko^c,
Irene Wagner-Döbler^d and Brigitte Kunze^d

^aEnvironmental and Computational Chemistry Group, Faculty of Pharmaceutical Sciences, University of Cartagena, Cartagena, Colombia; ^bAgrochemical Research Group, Chemistry Program, Faculty of Exact and Natural Sciences, University of Cartagena, Cartagena, Colombia; ^cChromatography Laboratory, Research Centre for Biomolecules, CIBIMOL, CENIVAM, Industrial University of Santander, Bucaramanga, Colombia; ^dHelmholtz Centre for Infection Research, Group Microbial Communication, Braunschweig, Germany

(Received 26 June 2010; final version received 20 January 2011)

Essential oils from Colombian plants were characterised by GC–MS, and assayed for anti-quorum sensing activity in bacteria sensor strains. Two major chemotypes were found for *Lippia alba*, the limonene-carvone and the citral (geranial-neral). For other species, the main components included α -pinene (*Ocotea* sp.), β -pinene (*Swinglea glutinosa*), cineol (*Elettaria cardamomum*), α -zingiberene (*Zingiber officinale*) and pulegone (*Minthostachys mollis*). Several essential oils presented promising inhibitory properties for the short chain AHL quorum sensing (QS) system, in *Escherichia coli* containing the biosensor plasmid pJBA132, in particular *Lippia alba*. Moderate activity as anti-QS using the same plasmid, were also found for selected constituents of essential oils studied here, such as citral, carvone and α -pinene, although solely at the highest tested concentration ($250 \mu\text{g mL}^{-1}$). Only citral presented some activity for the long chain AHL QS system, in *Pseudomonas putida* containing the plasmid pRK-C12. In short, essential oils from Colombian flora have promising properties as QS modulators.

Keywords: essential oil; Colombian plants; quorum sensing

1. Introduction

Essential oils are complex mixtures of volatile compounds isolated from plant materials. They are secondary metabolites chemically derived from terpenes and their oxygenated compounds, and each component contributes to their biological activities. These complex mixtures have different pharmacological properties (Firuzi, Asadollahi, Gholami, & Javidnia, 2010), including anti-inflammatory (Tavares et al., 2010), anti-tubercular (Bueno, Martínez, Stashenko, & Ribon, 2009), antibacterial (Okoh, Sadimenko, & Afolayan, 2010), antifungal (Hashem, Moharam, Zaied, & Saleh, 2010), protective of the gastrointestinal tract (Wang et al., 2008),

*Corresponding author. Email: joliverov@unicartagena.edu.co

anticonvulsant (Quintans-Júnior, Souza, Leite, Lessa, & Bonjardim, 2008; Viana, do Vale, Silva, & Matos, 2000) and antioxidant properties (Loizzo et al., 2009; Singh et al., 2010).

The antipathogenic properties of plants have received much less attention, but may be just as important in combating disease as their antibacterial counterparts (Adonizio, Downum, Bennett, & Mathee, 2006). The interruption of quorum sensing (QS), or bacterial cell-to-cell communication, is one example of an antipathogenic effect. Since a large number of systems affecting pathogenicity are controlled by QS, interrupting this communication system can render pathogenic bacteria non-virulent (Zhang & Dong, 2004). Inappropriate use or overuse of antibiotics has led to an increase in antibiotic resistance that has a great impact on human health around the world. The need for new antibiotics with mechanisms of action different from traditional ones is fundamental to increase survival probabilities in clinical infections where resistance is global. One of the existing alternatives to finding new antibacterial compounds relies on a sophisticated bacterial communication process called 'QS' (Raina et al., 2009). This process is important for bacteria to regulate community-wide behaviours including biofilm formation, virulence, conjugation, sporulation and swarming motility (Jayaraman & Wood, 2008).

QS allows bacteria to monitor its cell density through the release of specific signalling molecules called autoinducers. During growth, these chemicals are accumulated in the local environment until reaching a threshold concentration that initiates the signalling required for changes in gene expression, many of them involved in the regulation of a battery of virulence genes (Lynch & Wiener-Kronish, 2008). Gram-negative bacteria use N-acyl homoserine lactones (AHLs) as autoinducers, whereas Gram-positive bacteria rely on peptides, which after binding to specific receptors, initiate the signal cascade leading to the activation of systems that help these cells to succeed during host infection (Li et al., 2007). Based on this novel mechanism of action, QS has been suggested as an opportunity to fight bacterial infection/virulence by means other than growth inhibition, overcoming the problem linked to antibiotic resistance (Hentzer & Givskov, 2003; Lynch & Wiener-Kronish, 2008). Currently, many compounds, both synthetic and natural have been reported to have an anti-QS activity. The first group includes methylthioadenosine analogs (Gutierrez et al. 2009), whereas furanones (Lönn-Stensrud, Landin, Benneche, Petersen, & Scheie, 2009), cinnamaldehyde and substituted cinnamaldehyde (Brackman et al., 2008), are among the natural ones.

Despite the fact that Colombia has an immense biodiversity, few studies have been reported on the biological properties of essential oils from plants grown in this country. Among those, species such as *Lippia alba*, *Ocotea* sp, *Elettaria cardamomum*, *Swinglea glutinosa*, *Myntotachys mollis* and *Zingiber officinale* have been found to possess several biological properties, such as antioxidant, antifungal, antimicrobial, insect repellent, antiprotozoal, etc. (Bueno et al., 2009; Jaramillo, Stashenko, & Martínez, 2010; Olivero, Gonzalez, Güette, Jaramillo, & Stashenko, 2010; Stashenko, Jaramillo, & Martínez, 2004a; Weniger et al., 2001). In this work, we investigated if essential oils from plants grown in Colombia might have a potential as anti-QS substances, and therefore tested their ability to inhibit AHL-based QS.

Table 1. Essential oils from Colombian plants used in this study.

Species	Family	Sampling site	Extraction method	Voucher
<i>L. alba</i> (1) ^a	<i>Verbenaceae</i>	Bolívar	MWHD	512272
<i>S. glutinosa</i>	<i>Rutaceae</i>	Arauca	MWHD	521530
<i>L. alba</i> (2)	<i>Verbenaceae</i>	Cundinamarca	HD	484350
<i>L. alba</i> (3)	<i>Verbenaceae</i>	Tolima	MWHD	484350
<i>L. alba</i> (4)	<i>Verbenaceae</i>	Tolima	HD	484350
<i>M. mollis</i>	<i>Labiatae</i>	Cundinamarca	HD	521089
<i>L. alba</i> (5)	<i>Verbenaceae</i>	Arauca	MWHD	512084
<i>Ocotea</i> sp	<i>Lauraceae</i>	Boyacá	MWHD	CENIVAM-457 ^b
<i>E. cardamomum</i>	<i>Zingiberaceae</i>	Antioquia	HD	CENIVAM-450 ^b
<i>Z. officinale</i>	<i>Zingiberaceae</i>	Caldas	HD	CENIVAM 453 ^b

Notes: ^a Numbers in parenthesis refer to *L. alba* essential oil sampled from a different location or extracted using a specific method. ^bSamples with vouchers named CENIVAM are stored in our herbarium located in Bucaramanga (Colombia).

HD: hydrodistillation; MWHD: microwave assisted hydrodistillation.

2. Results and discussion

2.1. Essential oil composition

Table 1 shows the essential oils from Colombian plants used in this study. It presents scientific names of the plants, family, geographic site of collection, extraction method and identification number of the species. The major chemical components of the essential oils tested in this study are shown in Figure 1. In the case of *L. alba*, two different chemotypes were found, the limonene-carvone and the citral (geranial-neral). α -pinene and β -pinene were found to be the most abundant components in *Ocotea* sp. and *S. glutinosa*, respectively; whereas cineol, α -zingiberene and pulegone for *E. cardamomum*, *Z. officinale* and *M. mollis*, respectively. Large variations have been observed in the volatile composition of essential oils, depending on the part of the plant employed in the distillation, plant's state of development, geographic location, characteristics of the soil, climate and other local conditions (Stashenko et al., 2004a).

It is clear that the composition of the essential oil of *L. alba* depends on the collection site and the corresponding chemotype. The main constituents of *L. alba* (collected in Cundinamarca, Tolima and Arauca departments) were carvone and limonene. However, in *L. alba* (Collected in Bolívar Department), the main components were geranial and neral (Figure 1). Although quantitative differences are verified, the composition of the essential oils of *L. alba* shows agreement with the existence of different chemotypes as previously described (Bounatirou et al., 2007; Oliveira et al., 2006; Zoghbi, Andrade, Santos, Silva, & Maia, 1998). The principal components of *Ocotea* sp. were α -pinene, *p*-cymene and β -pinene. This composition is different from that found for *Ocotea quixos* (Ishpingo) from Ecuador, oil that has *trans*-cinnamaldehyde and methyl cinnamate as major constituents (Bruni et al., 2004). More than 350 species belong to the *Ocotea* genus, mainly distributed in the Americas and in Southern Africa, and those are widely known as commercial sources of spicy essential oils.

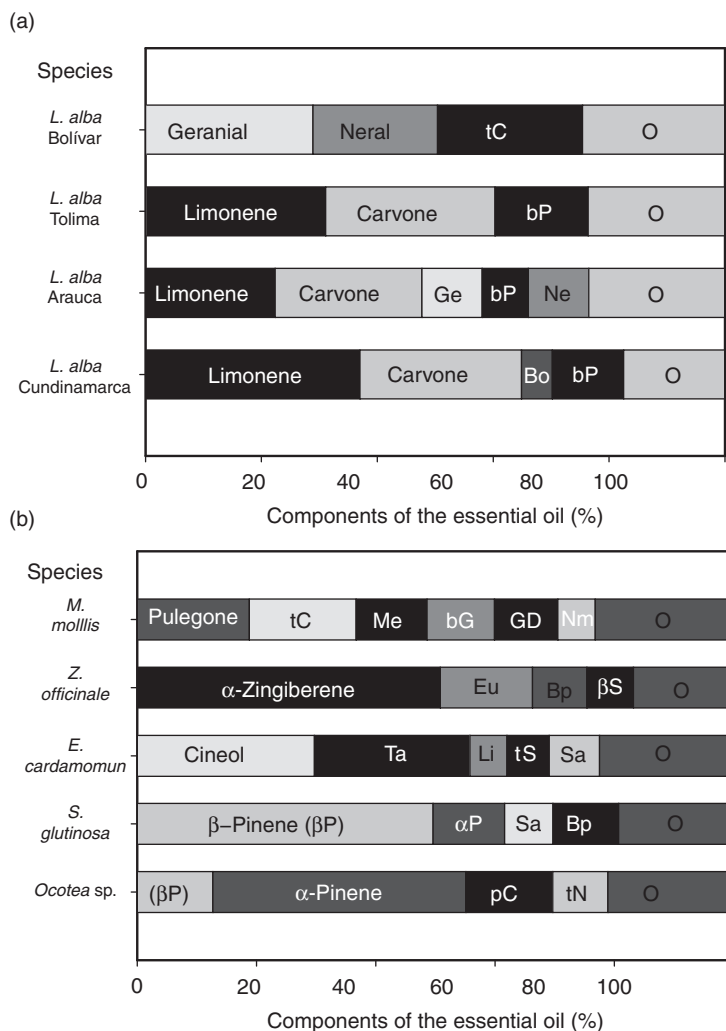


Figure 1. Composition of essential oils from different plants collected in Colombia. (a) Genus *Lippia*. (b) Other genus. Abbreviations. α P, α -pinene; β P, β -pinene; β S, β -sesquiphellandrene; pC, *p*-cymene; tC, *trans*-caryophyllene; tN, *trans*-nerolidol; bG, bicyclogermacrene; bP, bicyclosesquiphellandrene; Bo, bourbone; Ci, 1,8-cineol; Eu, eugenol; Ge, geraniol; GD, germacrene D; Me, menthol; Nm, neomenthol; Sa, sabinene; Li, linalool; Ne, neral; tS, *trans*-acetate sabinile hydrate; Ta, α -terpinyl acetate; O, other components.

The main components found in the volatile oil from *E. cardamomum* were 1,8-cineol and α -terpineol acetate. In contrast, Marongiu, Piras, and Porcedda. (2004) found as majority compounds in *E. cardamomum* α -terpinyl acetate, 1,8-cineole, linalyl acetate, limonene and linalool. Regarding the other studied species, major constituents for Colombian *M. mollis* were pulegone, menthone and *trans*- β -caryophyllene. Alkire, Tucker, and Maciarelo (1994) reported the major compounds obtained in the essential oil from ecuatorial *M. mollis*: neomenthol, menthone, menthol and piperitone.

Table 2. Anti-QS of essential oils isolated from Colombian plants.

Species	Anti-QS activity (% inhibition after 24 h exposure ^a)	
	<i>P. putida</i> (pKR-C12)	<i>E. coli</i> (pJBA132)
<i>L. alba</i> (1)	65	18
<i>S. glutinosa</i>	Null	28–73
<i>L. alba</i> (2)	Null	15–84
<i>L. alba</i> (3)	1	72–78
<i>L. alba</i> (4)	Null	61–62
<i>M. mollis</i>	Null	56–65
<i>L. alba</i> (5)	9	44–72
<i>Ocotea</i> sp.	25	71–76
<i>E. cardamomum</i>	31	21–22
<i>Z. officinale</i>	Null	34–69

Notes: ^aTested concentration was 1.2 mg mL⁻¹. For numbers in parenthesis refer to *L. alba* essential oil sampled from a different location or extracted using a specific method (see Table 1).

For Colombian essential oil from *S. glutinosa* the major components found were: β -pinene, α -pinene sabinene and bicyclosesquiphellandrene. Whereas (E)-nerolidol and caryophyllene oxide were the major components found in the essential oil from Cuban *S. glutinosa* (Pino, Marbot, & Fuentes, 2006). And for Colombian *Z. officinale* were: α -zingiberene, eugenol, bicyclosesquiphellandrene. In comparison, the essential oils obtained by hydrodistillation of the leaves and rhizomes of *Z. officinale* from Malaysia, the leaf oil was clearly dominated by β -caryophyllene, while the oil from the rhizomes was predominantly monoterpenoid, with camphene, geranial and geranyl acetate the three most abundant constituents (Sivasothy et al., 2011).

2.2. Inhibition of QS

Anti-QS properties for examined essential oils are shown in Table 2. Differences in activity for essential oils of *L. alba* are clearly dependent on the extraction method and the region where collection took place (Stashenko et al., 2004a; Zoghbi et al., 1998). Most tested oils had anti-QS properties for the sensor plasmid pJBA132, but few were active using the long-chain sensor plasmid pRK-C12. Essential oils of *L. alba* and *Ocotea* sp. were the most active.

Inhibition of QS for different selected constituents of essential oils are presented in Figure 2. Citral, at the highest tested concentration (250 μ M), was the only essential oil constituent that presented some activity as a QS inhibitor for the long-chain sensor plasmid pKR-C12 (*P. putida*). However, all examined compounds elicited moderate to good inhibition when examined against the sensor plasmid pJBA132 (*E. coli*). This is not surprising considering that pKR-C12 responds to long-chain AHLs (C12), whereas pJBA132 (*E. coli*) sensors short-chain AHLs (C6) (Wagner-Döbler et al., 2005). Although geranyl acetate has a total number of carbon atoms of 12 (C₁₂H₂₀O₂), it does not fit the 'long-chain' requirement because it is a

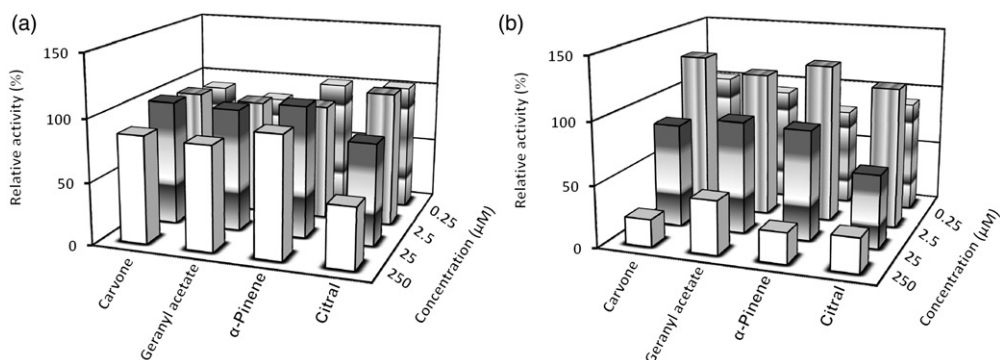


Figure 2. Anti-QS activity of different constituents of essential oils, measured as % activity relative to AHL (100%). QS in (a) *P. putida* (pKR-C12), and (b) *E. coli* (pJBA132).

branched molecule, as the other ones, and this probably prevents it from reaching the appropriate molecular size for binding on the receptor. On the other hand, both geranial and neral (citral) are linear compounds that seem to have some interference with the signalling induced by C₁₂-HSL; however, the mechanisms involved in the observed effects are still unknown.

The search for new chemicals that interfere with QS-related mechanisms in bacteria has considerably increased during the last years, opening up an alternative strategy for antimicrobial chemotherapy through the discovery of safer and less antibiotic resistant anti-bacterial drugs (Vattem, Mihalik, Crixell, & McLean, 2007). As a consequence, compounds from plants (Adonizio et al., 2006; Limsuwan, & Voravuthikunchai, 2008), animals (Park, Kaufmann, Bowen, Arbiser, & Janda, 2008), as well as from computational methods such as virtual screening (Kiran, Adikesavan, Cirioni, Giacometti, & Silvestri, 2008), have been found to have good anti-QS properties. Szabó, Varga, Hohmann, Schelz, and Szegedi. (2010) conducted studies about inhibition of QS signals by essential oils, showing that rose, geranium, lavender and rosemary oils were the most potent QS inhibitors. Eucalyptus and citrus oils moderately reduced pigment production by CV026, whereas the chamomile, orange and juniper oils were ineffective. Adonizio et al. (2006) worked with 50 medicinal plants from southern Florida which were screened for anti-QS activity using two biomonitor strains, *Chromobacterium violaceum* and *Agrobacterium tumefaciens*. Of these plants, six showed QS inhibition: *Conocarpus erectus* L. (Combretaceae), *Chamaecyfe hypericifolia* (L.) Millsp. (Euphorbiaceae), *Callistemon viminalis* (Sol. ex Gaertn.) G. Don (Myrtaceae), *Bucida burceras* L. (Combretaceae), *Tetrazygia bicolor* (Mill.) Cogn. (Melastomataceae), and *Quercus virginiana* Mill. (Fagaceae).

The results presented here have shown that essential oils are a diverse and suitable source of anti-QS molecules. The presence of molecules with different structures in the oils, also allows a wider spectrum of bioactivity. This is clearly seen with the two *L. alba* chemotypes studied here. The limonene-carvone chemotype has a greater activity against QS mediated by 3-oxo-C₆-HSL and no activity against the

system activated by C₁₂-HSL. In contrast, the citral chemotype (geranial-neral), has the opposite preference. Structurally speaking, limonene and carvone are not only cyclic monoterpenes but in terms of total length, they are more similar to a C₆ linear carbon skeleton, and consequently more likely to compete with a C₆-HSL. Moreover, as previously discussed, pKR-C12 has sensitivity for compounds with C₁₂-HSL, although this almost disappears acyl chains shorter than C8 (Steidle et al., 2002), explaining the lack of activity of the *L. alba* limonene/carvone chemotype in the *P. putida* strain.

Although anti-QS properties have not been reported for major components found in assayed essential oils, some of these compounds have been shown to be promising antimicrobial agents. Carvone, for instance, has also been identified as a major constituent in oils with antibacterial properties against both gram positive and gram negative bacteria (Shahat et al., 2008). Moreover, this terpenoid can act synergistically with other antibacterial compounds. Carvone alone, or the essential oil of *Mentha spicata* and *Anethum graveolens*, containing 40.1 and 20.3% carvone, augmented the nitrofurantoin activity against *Enterobacter cloacae* (Rafii & Shahverdi, 2007). As presented here, *Lippia alba* components are recognised as an important source of antimicrobial agents (Hennebelle, Sahpaz, Joseph, & Bailleul, 2008; Oliveira et al., 2006). In fact, not only this species, but in general others belonging to the same genus, have been used in traditional herbal medicine to treat several respiratory problems, including colds and coughs (Viljoen, Subramoney, van Vuuren, Başer, & Demirci, 2005). Despite the fact that biological activity of essential oils is usually linked to the presence of major components (Firuzi et al., 2010), in some cases, those chemicals present in low concentrations could also be responsible for the observed biological activities (Grassmann, Hippeli, Vollmann, & Elstner, 2003). Regarding species different from *Lippia*, other compounds detected in essential oils are also of special interest. It has been reported that pulegone, thymol or cineol-rich essential oils have also been effective against Gram-positive and Gram-negative bacteria (Oumzil et al., 2002; Shapira, & Mimran, 2007).

Despite the good anti-QS properties observed for tested essential oils, reported toxicity, based on the brine shrimp mortality method, suggests that most of the evaluated essential oils were cytotoxic, with LC₅₀ values closer or below 20 µg mL⁻¹ (Olivero et al., 2010). Accordingly, additional pharmacological actions for these oils should also be considered. Several investigators have reported the use of *L. alba* as neurosedative (Hennebelle et al., 2008), anticonvulsivant (Viana et al., 2000), antioxidant (Stashenko et al., 2004a) and antiviral (Andrighetti-Fröhner et al., 2005), among others.

In short, results presented here suggest that essential oils from Colombian flora have interesting anti-QS activities which may make them targets for the development of antimicrobial drugs.

3. Experimental

3.1. Bacterial strains

The sensor strains were grown at 37°C on LB medium with 25 µg mL⁻¹ tetracycline (*E. coli* MT102/pJBA132) and 20 µg mL⁻¹ gentamycin (*Pseudomonas putida* F117/pKRC12), respectively.

3.2. Collection of plant materials and essential oil extraction

Plant specimens were collected from several locations along Colombia. Sampling sites and extraction method are shown in Table 1. Samples were stored in the dark and transported to the laboratory, washed and used fresh with minimal light exposure. Plant material was identified by botanist Jaime Aguirre Santoro from the Institute of Natural Sciences at the National University of Colombia (Bogotá). A voucher of each species is deposited at the Institute's Herbarium.

3.3. Extractions

Essential oils were extracted by hydrodistillation (HD) or microwave assisted hydrodistillation (MWHd). HD was performed in a 5 L-round flask with 500 g of plant material and 4 L of water, using an electric heater (boiling water) for 2 h. The oil, previously saturated with NaCl, was decanted from the condensate and dried with anhydrous sodium sulfate (Stashenko et al., 2004a; Stashenko, Jaramillo, & Martinez, 2004b). MWHd consisted of a hydrodistillation apparatus placed inside a domestic microwave oven (Kendo, 2.45 GHz, 800 W), with a side orifice through which an external glass condenser joined the round flask with the plant material (100 g) and water (2 L), inside the oven. The oven was operated for 30 min at full power, which caused water to boil vigorously and reflux (Paré, Bélanger, & Stafford, 1994; Stashenko et al., 2004a; Stashenko et al., 2004b).

3.4. GC-MS Analysis

The essential oil analysis was carried out using an *Agilent Technologies* 6890 Plus equipped with a HP-5 MS capillary column (30 m $\times$ 0.25 mm i.d. $\times$ 0.25 μ m, d_f), coupled to an *Agilent Technologies* MSD 5973 mass-selective detector. For GC-MS detection, an electron ionisation system with ionisation energy of 70 eV was used. Helium was the carrier gas at a flow rate of 1 mL min⁻¹. Diluted samples (1:100 v/v, in CH₂Cl₂) of 1.0 μ L were injected manually and in the *splitless* mode. Components were identified on the basis of comparison of their relative retention time and mass spectra with those of some standard compounds, NBS75K library data of the GC-MS system, and reported data (Adams, 1995; Davies, 1990).

3.5. QS inhibition assay

Inhibition of QS systems driven by AHLs was evaluated using biosensor strains that do not produce AHLs by their own but are capable of sensing their presence through a sensor plasmid which expresses an AHL controlled promoter fused to a gene coding for fluorescence (gfp). *P. putida* F117 (pKR-C12) was used to detect inhibitors of long-chain AHLs (C12), and *E. coli* MT102 (pJBA132) was used for the detection of short chain AHL (C6) inhibitors (Wagner-Döbler et al., 2005). In short, overnight cultures (50 mL) of these strains were diluted with the same volume of accordant fresh medium and incubated for one more hour. These cultures adjusted to an OD₆₂₀ of 1.0 were used for the bioassay. Aliquots of 100 μ L were incubated at 30°C in microtiter plates together with 90 μ L of the medium, 5 μ L of the essential oil (0.025–1.25 mg mL⁻¹) or essential oil's component, and 5 μ L of the corresponding

AHL ($0.25 \mu\text{g mL}^{-1}$). Samples containing only the corresponding AHL without test compound were used as positive control and samples containing only solvent ($10 \mu\text{L}$) with neither test compound nor AHL were used as negative control. After 24 h, fluorescence at an excitation wavelength of 485 nm and a detection wavelength of 535 nm, and growth at an OD_{620} was determined in a Victor 1420 Multilabel Counter. Afterwards the fold induction of fluorescence in the test samples was calculated by dividing their specific fluorescence ($\text{gfp}_{535}/\text{OD}_{620}$) by the specific fluorescence of the negative control. Percent of inhibition was calculated by comparing the fold induction of positive control samples (only AHL) to samples with AHL and essential oils. Anti-QS in one experiment was measured in triplicate and data are presented for at least two independent experiments.

Due to the complex and diverse nature of components present in the essential oils, several individual compounds and an isomeric mixture (citral, geranial + neral) that were found both as major components (R-carvone 98%, citral 95%, from Aldrich Chemical Corp., Milwaukee, WI, USA; and α -pinene 98.5% from Fluka,) as well as a minor constituent (geranyl acetate 99%, from Fluka) were also tested for QS inhibition.

4. Conclusions

In short, results presented here suggest that essential oils from Colombian flora have interesting anti-QS activities which may make them targets for the development of antimicrobial drugs.

Acknowledgements

The authors thank Colciencias, Bogota, Colombia (Grant no. RC 432-2004), and the Von Humboldt Alexander Foundation (Germany). The authors also thank Birte Engelhardt for skillfull assistance and Leo Eberl for providing the biosensor strains for the QS tests.

References

- Adams, R.P. (1995). *Identification of essential oil components by gas chromatography/mass spectrometry*. Carol Stream, IL: Allured Publishing Corporation.
- Adonizio, A.L., Downum, K., Bennett, B.C., & Mathee, K. (2006). Anti-quorum sensing activity of medicinal plants in southern Florida. *Journal of Ethnopharmacology*, 105, 427–435.
- Alkire, B.H., Tucker, A.O., & Maciarello, M.J. (1994). Tipo, *minthostachys mollis* (lamiaceae): an ecuadorian mint. *Economic Botany*, 48, 60–64.
- Andrighetti-Fröhner, C.R., Sincero, T.C., da Silva, A.C., Savi, L.A., Gaido, C.M., Bettega, J.M., . . . , Simões, C.M. (2005). Antiviral evaluation of plants from Brazilian Atlantic Tropical Forest. *Fitoterapia*, 76, 374–378.
- Brackman, G., Defoirdt, T., Miyamoto, C., Bossier, P., Van Calenbergh, S., Nelis, H., & Coenye, T. (2008). Cinnamaldehyde and cinnamaldehyde derivatives reduce virulence in *Vibrio* spp. by decreasing the DNA-binding activity of the quorum sensing response regulator LuxR. *BMC Microbiology*, 8, 149.
- Bruni, R., Medici, A., Andreotti, E., Fantin, C., Muzzoli, M., Dehesa, M., . . . , Sacchetti, G. (2004). Chemical composition and biological activities of Ishpingo essential oil, a

- traditional Ecuadorian spice from *Ocotea quixos* (Lam.) Kosterm. (Lauraceae) flower calices. *Food Chemistry*, 85, 415–421.
- Bounatirou, S., Smiti, S., Miguel, M.G., Faleiro, L., Rejeb, M.N., Neffati, M., . . . , Pedro, L.G. (2007). Chemical composition, antioxidant and antibacterial activities of the essential oils isolated from Tunisian *Thymus capitatus* Hoff. et Link. *Food Chemistry*, 105, 146–155.
- Bueno, J.G., Martínez, J.R., Stashenko, E., & Ribon, W. (2009). Antitubercular activity of eleven aromatic and medicinal plants occurring in Colombia. *Biomédica*, 29, 51–60.
- Davies, N.W. (1990). Gas chromatographic retention indices of monoterpenes and sesquiterpenes on methyl silicone and Carbowax 20M phases. *Journal of Chromatography*, 503, 1–24.
- Firuzi, O., Asadollahi, M., Gholami, M., & Javidnia, K. (2010). Composition and biological activities of essential oils from four *Heracleum* species. *Food Chemistry*, 122, 117–122.
- Gutierrez, J.A., Crowder, T., Rinaldo-Matthis, A., Ho, M.C., Almo, S.C., & Schramm, V.L. (2009). Transition state analogs of 5'-methylthioadenosine nucleosidase disrupt quorum sensing. *Nature Chemical Biology*, 5, 251–257.
- Grassmann, J., Hippeli, S., Vollmann, R., & Elstner, E.F. (2003). Antioxidative properties of the essential oil from *Pinus mugo*. *Journal of Agricultural and Food Chemistry*, 51, 7576–7582.
- Hashem, H., Moharam, A.M., Zaied, A.A., & Saleh, F.E.M. (2010). Efficacy of essential oils in the control of cumin root rot disease caused by *Fusarium* spp. *Crop Protection*, 29, 1111–1117.
- Hennebelle, T., Sahpaz, S., Joseph, H., & Bailleul, F. (2008). Ethnopharmacology of *Lippia alba*. *Journal of Ethnopharmacology*, 116, 211–222.
- Hentzer, M., & Givskov, M. (2003). Pharmacological inhibition of quorum sensing for the treatment of chronic bacterial infections. *The Journal of Clinical Investigation*, 112, 1300–1307.
- Jaramillo, B.E., Stashenko, E., & Martínez, J.R. (2010). Composición química volátil de *Satureja brownii* (Sw.) Briq. colombiana y determinación de su actividad antioxidante. *Revista Cubana de Plantas Medicinales*, 15, 52–63.
- Jayaraman, A., & Wood, T.K. (2008). Bacterial quorum sensing: Signals, circuits, and implications for biofilms and disease. *Annual Review of Biomedical Engineering*, 10, 145–167.
- Kiran, M.D., Adikesavan, N.V., Cirioni, O., Giacometti, A., & Silvestri, C. (2008). Discovery of a quorum-sensing inhibitor of drug-resistant staphylococcal infections by structure-based virtual screening. *Molecular Pharmacology*, 73, 1578–1586.
- Li, M., Lai, Y., Villaruz, A.E., Cha, D.J., Sturdevant, D.E., & Otto, M. (2007). Gram-positive three-component antimicrobial peptide-sensing system. *Proceedings of the National Academy of Sciences USA*, 104, 9469–9474.
- Limsuwan, S., & Voravuthikunchai, S.P. (2008). *Boesenbergia pandurata* (Roxb.) Schltr., *Eleutherine americana* Merr. and *Rhodomyrtus tomentosa* (Aiton) Hassk. as antibiofilm producing and anti-quorum sensing in *Streptococcus pyogenes*. *FEMS Immunology and Medical Microbiology*, 53, 429–436.
- Loizzo, M.R., Menichini, F., Conforti, F., Tundis, R., Bonesi, M., Saab, A., . . . , Frega, N. (2009). Chemical analysis, antioxidant, anti-inflammatory and anticholinesterase activities of *Origanum ehrenbergii* Boiss and *Origanum syriacum* L. essential oils. *Food Chemistry*, 117, 174–180.
- Lönn-Stensrud, J., Landin, M.A., Benneche, T., Petersen, F.C., & Scheie, A.A. (2009). Furanones, potential agents for preventing *Staphylococcus epidermidis* biofilm infections? *Journal of Antimicrobial Chemotherapy*, 63, 309–316.

- Lynch, S.V., & Wiener-Kronish, J.P. (2008). Novel strategies to combat bacterial virulence. *Current Opinion in Critical Care*, 14, 593–599.
- Marongiu, B., Piras, A., & Porcedda, S. (2004). Comparative analysis of the oil and supercritical CO₂ extract of *Elettaria cardamomum* (L.) Maton. *Journal of Agricultural Food Chemistry*, 52, 6278–6282.
- Okoh, O.O., Sadimenko, A.P., & Afolayan, A.J. (2010). Comparative evaluation of the antibacterial activities of the essential oils of *Rosmarinus officinalis* L. obtained by hydrodistillation and solvent free microwave extraction methods. *Food Chemistry*, 120, 308–312.
- Oliveira, D.R., Leitão, G.G., Santos, S.S., Bizzo, H.R., Lopes, D., Alviano, C.S., . . . , Leitão, S.G. (2006). Ethnopharmacological study of two *Lippia* species from Oriximiná, Brazil. *Journal of Ethnopharmacology*, 108, 103–108.
- Olivero, J., Gonzalez, T., Güette, J., Jaramillo, B., & Stashenko, E. (2010). Chemical composition and antioxidant activity of essential oils isolated from Colombian plants. *Brazilian Journal of Pharmacognosy*, 20, 568–574.
- Oumzil, H., Ghoulami, S., Rhajaoui, M., Iidirissi, A., Fkih-Tetouani, S., Faid, M., & Benjouad, A. (2002). Antibacterial and antifungal activity of essential oils of *Mentha suaveolens*. *Phytotherapy Research*, 16, 727–731.
- Paré, J.R., Bélanger, J.M.R., & Stafford, S.S. (1994). Microwave-Assisted process (MAPTM): A new tool for the analytical laboratory. *Trends in Analytical Chemistry*, 13, 176–184.
- Park, J., Kaufmann, G.F., Bowen, J.P., Arbiser, J.L., & Janda, K.D. (2008). Solenopsin A, a venom alkaloid from the fire ant *Solenopsis invicta*, inhibits quorum-sensing signaling in *Pseudomonas aeruginosa*. *The Journal of Infectious Diseases*, 198, 1198–1201.
- Pino, J.A., Marbot, R., & Fuentes, V. (2006). Aromatic Plants from Western Cuba IV. Composition of the Leaf Oils of *Clausena lansium* (Lour.) Skeels and *Swinglea glutinosa* (Blanco) Merr. *Journal of Essential Oil Research*, 18, 139–141.
- Quintans-Júnior, L.J., Souza, T.T., Leite, B.S., Lessa, N.M.N., Bonjardim, L.R., & Santos, M.R.V. (2008). Phytochemical screening and anticonvulsant activity of *Cymbopogon winterianus* Jowitt (Poaceae) leaf essential oil in rodents. *Phytomedicine*, 15, 619–624.
- Rafii, F., & Shahverdi, A.R. (2007). Comparison of essential oils from three plants for enhancement of antimicrobial activity of nitrofurantoin against enterobacteria. *Chemotherapy*, 53, 21–25.
- Raina, S., De Vizio, D., Odell, M., Clements, M., Vanhulle, S., & Keshavarz, T. (2009). Microbial quorum sensing: A tool or a target for antimicrobial therapy? *Biotechnology and Applied Biochemistry*, 54, 65–84.
- Shahat, A.A., El-Barouty, G., Hassan, R.A., Hammouda, F.M., Abdel-Rahman, F.H., & Saleh, M.A. (2008). Chemical composition and antimicrobial activities of the essential oil from the seeds of *Enterolobium contortisiliquum* (Leguminosae). *Journal of Environmental Science and Health, Part B*, 43, 519–525.
- Shapira, R., & Mimran, E. (2007). Isolation and characterization of *Escherichia coli* mutants exhibiting altered response to thymol. *Microbial Drug Resistance. Fall*, 13, 157–65.
- Singh, P., Shukla, R., Prakash, B., Kumar, A., Singh, S., Mishra, P.K., & Dubey, N.K. (2010). Chemical profile, antifungal, antiaflatoxigenic and antioxidant activity of *Citrus maxima* Burm. and *Citrus sinensis* (L.) Osbeck essential oils and their cyclic monoterpene, dl-limonene. *Food and Chemical Toxicology*, 48, 1734–1740.
- Sivasothy, Y., Chong, W.K., Hamid, A., Eldeen, I.M., Sulaiman, S.F., & Awang, K. (2011). Essential oils of *Zingiber officinale* var. rubrum Theilade and their antibacterial activities. *Food Chemistry*, 124, 514–517.
- Stashenko, E., Jaramillo, B., & Martinez, J. (2004a). Comparison of different extraction methods for the analysis of volatile secondary metabolites of *Lippia alba* (Mill.)

- N.E. Brown, grown in Colombia, and evaluation of its in vitro antioxidant activity. *Journal of Chromatography A*, 1025, 93–103.
- Stashenko, E., Jaramillo, B., & Martínez, J. (2004b). Analysis of volatile secondary metabolites from Colombian *Xylopia aromatica* (Lamarck) by different extractive and headspace methods and gas chromatography. *Journal of Chromatography A*, 1025, 105–113.
- Steidle, A., Allesen-Holm, M., Riedel, K., Berg, G., Givskov, M., Molin, S., & Eberl, L. (2002). Identification and characterization of an N-acylhomoserine lactone-dependent quorum-sensing system in *Pseudomonas putida* strain IsoF. *Applied and Environmental Microbiology*, 68, 6371–6382.
- Szabó, M.A., Varga, G.Z., Hohmann, J., Schelz, Z., & Szegedi, E. (2010). Inhibition of quorum-sensing signals by essential oils. *Phytotherapy Research: PTR*, 24, 782–6.
- Tavares, A.C., Gonçalves, M.J., Cruz, M.T., Cavaleiro, C., Lopes, M.C., Canhoto, J., & Salgueiro, L.R. (2010). Essential oils from *Distichoselinum tenuifolium*: chemical composition, cytotoxicity, antifungal and anti-inflammatory properties. *Journal of Ethnopharmacology*, 130, 593–598.
- Vattem, D.A., Mihalik, K., Crixell, S.H., & McLean, R.J.C. (2007). Dietary phytochemicals as quorum sensing inhibitors. *Fitoterapia*, 78, 302–310.
- Viana, G.S., do Vale, T.G., Silva, C.M., & Matos, F.J. (2000). Anticonvulsant activity of essential oils and active principles from chemotypes of *Lippia alba* (Mill.) N.E. Brown. *Biological & Pharmaceutical Bulletin*, 23, 1314–1317.
- Viljoen, A.M., Subramoney, S., van Vuuren, S.F., Başer, K.H., & Demirci, B. (2005). The composition, geographical variation and antimicrobial activity of *Lippia javanica* (Verbenaceae) leaf essential oils. *Journal of Ethnopharmacology*, 96, 271–277.
- Wagner-Döbler, I., Thiel, V., Eberl, L., Allgaier, M., Bodor, A., Meyer, S., . . . , Schulz, S. (2005). Discovery of complex mixtures of novel long-chain quorum sensing signals in free-living and host-associated marine alphaproteobacteria. *Chembiochem*, 6, 2195–2206.
- Wang, X., Zhang, F., Liu, Z., Feng, H., Yu, Z.B., Lu, Y., . . . , Fan, D. (2008). Effects of essential oil from *Croton tiglium* L. on intestinal transit in mice. *Journal of Ethnopharmacology*, 117, 102–107.
- Weniger, B., Robledo, S., Arango, G.J., Deharo, E., Aragón, R., Muñoz, V., . . . , Anton, R. (2001). Antiprotozoal activities of Colombian plants. *Journal of Ethnopharmacology*, 78, 193–200.
- Zhang, L.H., & Dong, Y.H. (2004). Quorum sensing and signal interference: diverse implications. *Molecular Microbiology*, 53, 1563–1571.
- Zoghbi, M., Andrade, E., Santos, A., Silva, M., & Maia, J. (1998). Essential oils of *Lippia alba* (Mill.) N. E. Br growing wild in the Brazilian Amazon. *Flavour and Fragrance Journal*, 13, 47–48.